

## Two New Phenylpropanoid Derivatives from *Codonopsis tangshen* OLIV.

by Dan Song<sup>a) b) c)</sup>, Gui-Xin Chou<sup>\*a) c)</sup>, Guo-Yue Zhong<sup>b)</sup>, and Zheng-Tao Wang<sup>\*a) c)</sup>

<sup>a)</sup> Key Laboratory of Standardization of Chinese Medicines of Ministry of Education, Institute of Chinese Materia Medica, Shanghai University of Traditional Chinese Medicine, No. 1200 Cai Lun Road, Zhangjiang Hi-Tech Park, Shanghai, 201203, P. R. China

(phone: +86-21-51322507; fax: +86-21-51322519; e-mail: wangzht@hotmail.com)

<sup>b)</sup> Chongqing Academy of Chinese Materia Medica, Chongqing 400065, P. R. China

<sup>c)</sup> Shanghai R&D Centre for Standardization of Chinese Medicines, Shanghai, 201203, P. R. China

(phone: +86-21-51322507, +86-21-50271706; fax: +86-21-51322519, +86-21-50271708;

e-mail: wangzht@hotmail.com, chouguixin@hotmail.com)

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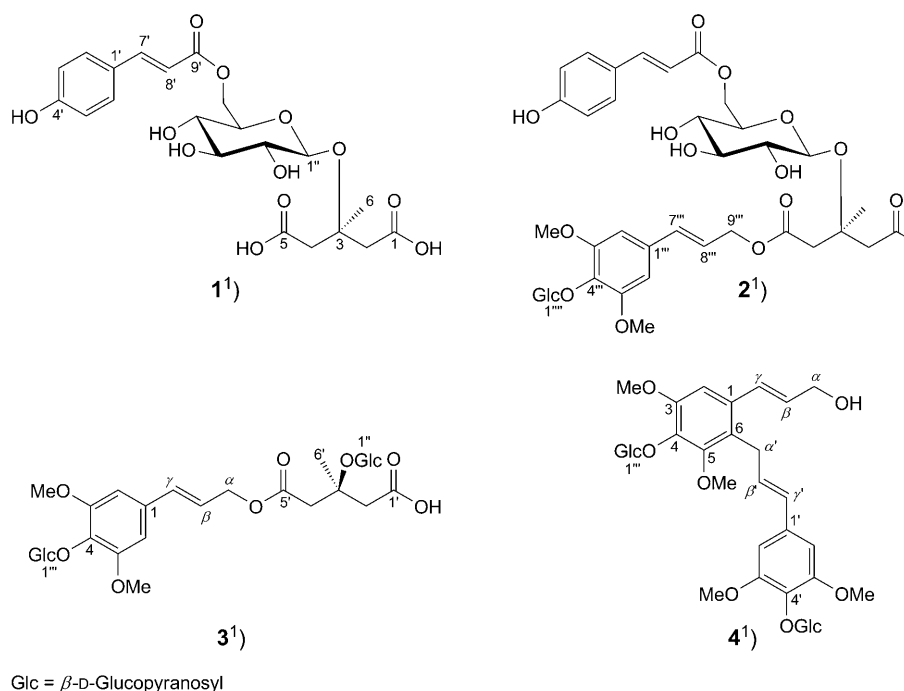
Two new 3-hydroxy-3-methylglutaric acid derived phenylpropanoid glucosides, tangshenoside V (**1**) and tangshenoside VI (**2**), were isolated from the roots of *Codonopsis tangshen* OLIV., along with the two known compounds tangshenoside I (**3**) and tangshenoside III (**4**). Their structures were elucidated by spectroscopic methods (IR and 1D- and 2D-NMR) and by mass spectrometry (HR-ESI-MS).

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**Introduction.** – The roots of *Codonopsis tangshen* OLIV. (Dangshen in Chinese) is a very common traditional Chinese medicine, belonging to the family Campanulaceae. It is recorded in the Chinese Pharmacopoeia for the treatment of neurosis, hematopoietic diseases, gastric ulcer, and nephritis [1][2]. Dangshen was also administered as a substitute for Asian ginseng, or as a tonic for immunoregulatory purposes [3][4]. Many compounds such as polyacetylenes, alkaloids, flavonoids, steroids, sesquiterpenes, triterpenoids, and polysaccharides have been isolated in the *Codonopsis* genus [5–9]. In previous works, four phenylpropanoid glucosides, tangshenoside I–IV, were isolated and identified from the roots of *C. tangshen* [6][7]. In the present paper, we describe the isolation and the structure elucidation of tangshenoside V (**1**) and tangshenoside VI (**2**), along with two derivatives previously isolated from this plant, tangshenosides I (**3**) and III (**4**).

**Results and Discussion.** – The air-dried and powdered roots were extracted with 70% EtOH to give the crude extract (10 kg). The total extract was suspended in H<sub>2</sub>O and partitioned successively with petroleum ether, AcOEt, and BuOH. The BuOH fraction was separated by column chromatography over *D-101* macroporous resin, silica gel, and *Sephadex LH-20*, repeatedly, followed by *ODS* column chromatography; it afforded a series of phenylpropanoid derivatives, including two new compounds **1** and **2** and the two known ones **3** and **4**. The structures of the known compounds were confirmed as tangshenoside I (**3**) and tangshenoside III (**4**) by comparison of their physical and spectral data with the reported data [7][8].

Compound **1**, a colorless gum, showed a molecular formula C<sub>21</sub>H<sub>26</sub>O<sub>12</sub> as deduced from its positive-mode HR-ESI-MS (*m/z* 493.1334 (C<sub>21</sub>H<sub>26</sub>NaO<sub>12</sub><sup>+</sup>)). The IR spectrum



revealed the absorption bands of OH ( $3432\text{ cm}^{-1}$ ) and CO ( $1706\text{ cm}^{-1}$ ) groups. From the  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR (*Table*), HMQC, and HMBC (*Fig. 1*) data, the structure of **1** was elucidated as 3-[(6-*O*-(*p*-coumaroyl)- $\beta$ -D-glucopyranosyl)oxy]-3-methylglutaric acid, named tangshenoside V. The configuration at the 3-hydroxy-3-methylglutaric acid derived moiety was not determined. However, it can be proposed as (3*S*), since all compounds containing such a residue reported until now in the family Campanulaceae possess this configuration [6–8].

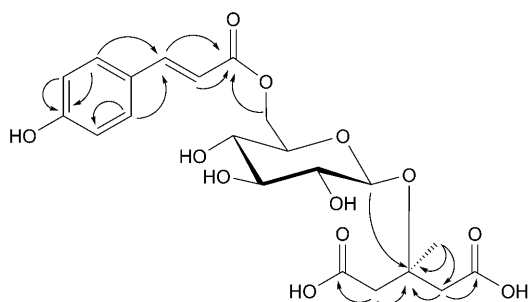


Fig. 1. Selected HMBC (H  $\rightarrow$  C) of **1**

<sup>1)</sup> Arbitrary atom numbering; for systematic names, see *Exper. Part*.

Table.  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR Data (500 and 125 MHz, resp.;  $\text{CD}_3\text{OD}$ ) of **1** and **2**<sup>1</sup>.  $\delta$  in ppm,  $J$  in Hz.

	<b>1</b>		<b>2</b>	
	$\delta(\text{H})$	$\delta(\text{C})$	$\delta(\text{H})$	$\delta(\text{C})$
C(1)		174.6		174.6
CH <sub>2</sub> (2)	2.80–2.92 ( <i>m</i> )	44.5	2.80–2.92 ( <i>m</i> )	44.5
C(3)		77.8		77.9
CH <sub>2</sub> (4)	2.80–2.92 ( <i>m</i> )	44.5	2.80–2.92 ( <i>m</i> )	44.7
C(5)		174.7		172.6
Me(6)	1.49 ( <i>s</i> )	25.0	1.50 ( <i>s</i> )	25.4
C(1')		127.5		127.5
H–C(2')	7.42 ( <i>d</i> , $J=8.6$ )	131.4	7.39 ( <i>d</i> , $J=8.2$ )	131.5
H–C(3')	6.78 ( <i>d</i> , $J=8.6$ )	117.1	6.78 ( <i>d</i> , $J=8.2$ )	117.1
C(4')		161.5		161.5
H–C(5')	6.78 ( <i>d</i> , $J=8.6$ )	117.1	6.78 ( <i>d</i> , $J=8.2$ )	117.1
H–C(6')	7.42 ( <i>d</i> , $J=8.6$ )	131.4	7.39 ( <i>d</i> , $J=8.2$ )	131.5
H–C(7')	7.62 ( <i>d</i> , $J=15.9$ )	147.0	7.60 ( <i>d</i> , $J=15.9$ )	147.0
H–C(8')	6.33 ( <i>d</i> , $J=15.9$ )	115.3	6.29 ( <i>d</i> , $J=15.9$ )	115.4
C(9')		169.3		169.3
H–C(1'')	4.65 ( <i>d</i> , $J=7.7$ )	98.6	4.65 ( <i>d</i> , $J=7.7$ )	98.7
H–C(2'')	3.22 ( <i>t</i> , $J=8.9$ )	75.3	3.22–3.49 ( <i>m</i> )	75.4
H–C(3'')	3.42 ( <i>t</i> , $J=8.9$ )	78.2	3.22–3.49 ( <i>m</i> )	78.1
H–C(4'')	3.30–3.34 ( <i>m</i> )	72.1	3.22–3.49 ( <i>m</i> )	72.2
H–C(5'')	3.53–3.56 ( <i>m</i> )	75.5	3.22–3.49 ( <i>m</i> )	75.5
CH <sub>2</sub> (6'')	4.48 ( <i>dd</i> , $J=11.8, 2.0$ ), 4.28 ( <i>dd</i> , $J=11.8, 6.8$ )	65.1	4.48 ( <i>dd</i> , $J=11.4, 2.0$ ), 4.28 ( <i>dd</i> , $J=11.4, 6.8$ )	65.1
C(1''')				134.9
H–C(2''')			6.71 ( <i>s</i> )	106.3
C(3''')				154.6
C(4''')				136.6
C(5''')				154.6
H–C(6''')			6.71 ( <i>s</i> )	106.3
MeO–C(3''')			3.83 ( <i>s</i> )	57.4
MeO–C(5''')			3.83 ( <i>s</i> )	57.4
H–C(7''')			6.56 ( <i>d</i> , $J=16.0$ )	135.2
H–C(8''')			6.23 ( <i>ddd</i> , $J=16.0, 6.5, 6.5$ )	124.6
CH <sub>2</sub> (9''')			4.65 ( <i>d</i> , $J=6.5$ )	66.3
H–C(1''''')			4.86 ( <i>d</i> , $J=7.4$ )	105.7
H–C(2''''')			3.22–3.49 ( <i>m</i> )	76.0
H–C(3''''')			3.22–3.49 ( <i>m</i> )	78.6
H–C(4''''')			3.22–3.49 ( <i>m</i> )	71.6
H–C(5''''')			3.22–3.49 ( <i>m</i> )	78.2
CH <sub>2</sub> (6''''')			3.79 ( <i>dd</i> , $J=12.0, 2.0$ ), 3.67 ( <i>dd</i> , $J=12.0, 4.9$ )	62.9

In the  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR spectra data of **1** (Table), three structural moieties were clearly observed. Two *d* at  $\delta(\text{H})$  6.78 (*d*,  $J=8.6$ , 2 H) and 7.42 (*d*,  $J=8.6$ , 2 H), two olefinic H-atoms at  $\delta(\text{H})$  6.33 (*d*,  $J=15.9$ , 1 H) and 7.62 (*d*,  $J=15.9$ , 1 H), together with four aromatic C-atoms, two olefinic C-atoms as well as one carboxy C-atom indicated the presence of a *p*-coumaroyl (= 3-(4-hydroxyphenyl)-1-oxoprop-2-en-1-yl) group. A Me *s* at  $\delta(\text{H})$  1.49 (*s*, Me(6)), and the four H-atoms at  $\delta(\text{H})$  2.80–2.92 (*m*, CH<sub>2</sub>(2), CH<sub>2</sub>(4)) corresponding to two CH<sub>2</sub> groups, together with two CO signals at  $\delta(\text{C})$  174.6 (C(1)) and 174.7

(C(5)) demonstrated the presence of a 3-hydroxy-3-methylglutaric acid moiety [5]. The anomeric signal at  $\delta(\text{H})$  4.65 (*d*,  $J = 7.7$ , 1 H), the corresponding  $^{13}\text{C}$ -NMR signal at  $\delta(\text{C})$  98.6, together with the signals in the region  $\delta(\text{H})$  3.22–4.48 and relevant  $^{13}\text{C}$ -NMR resonances, indicated the presence of a  $\beta$ -glucopyranosyl (Glc) unit. Interpretation of the HMQC and HMBC data of **1** (Fig. 1) revealed the substitution pattern and allowed us to assign all the  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR signals. The *p*-coumaroyloxy group was located at C(6'') of the Glc moiety because of the downfield shift observed for this H-atom in the  $^1\text{H}$ -NMR spectrum, as corroborated by the HMBC cross-peaks  $\text{CH}_2(6'')/\text{C}(9')$ . The attachment of the  $\beta$ -glucopyranose moiety at C(3) was established by the HMBC cross-peak  $\text{H}-\text{C}(1'')/\text{C}(3)$ .

Compound **2**, a colorless gum, had a molecular formula  $\text{C}_{38}\text{H}_{48}\text{O}_{20}$  as shown by its positive-mode HR-ESI-MS ( $m/z$  847.26298 ( $\text{C}_{38}\text{H}_{48}\text{NaO}_{20}^+$ )). The IR spectrum revealed the absorption bands of OH ( $3432\text{ cm}^{-1}$ ) and CO ( $1706\text{ cm}^{-1}$ ) groups. The UV spectrum of **2** showed absorptions at  $\lambda_{\text{max}}$  (MeOH) 227 and 277 nm, which suggested that there was an aromatic system conjugated with an unsaturated side chain. Comparison of the  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR spectra of **2** (Table) with those of **1** indicated that **2** is a monoesterified derivative of **1**. From these and the HMBC data (Fig. 2), the structure of **2** was elucidated as 3'-*O*-[6-*O*-(*p*-coumaroyl)- $\beta$ -D-glucopyranosyl]jussurienoside, named tangshenoside VI.

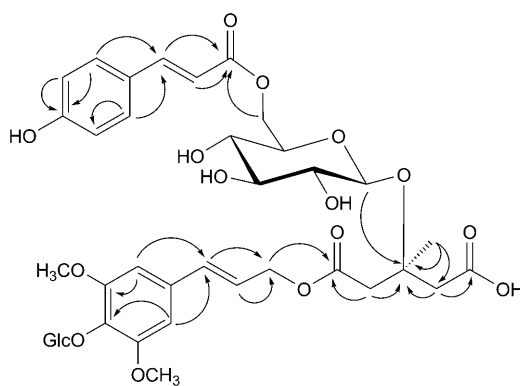


Fig. 2. Selected HMBC ( $\text{H} \rightarrow \text{C}$ ) of **2**

The presence of a (*2E*)-prop-2-en-1-ol derived moiety was indicated by a pair of olefinic signals at  $\delta(\text{H})$  6.56 (*d*,  $J = 16.0$ , 1 H) and 6.23 (*ddd*,  $J = 16.0, 6.5, 6.5$ , 1 H), and an oxygenated- $\text{CH}_2$  *d* at  $\delta(\text{H})$  4.65 (*d*,  $J = 6.5$ , 2 H). Furthermore, two aromatic H-atoms at  $\delta(\text{H})$  6.71 (*s*, 2 H) were attributed to a 1,3,4,5-tetrasubstituted phenyl ring. A six H-atoms *s* at  $\delta(\text{H})$  3.83 (*s*, 6 H) was characteristic of the two MeO groups symmetrically situated at a benzene ring. One *d* with a large coupling constant due to an anomeric H-atom at  $\delta(\text{H})$  4.86 (*d*,  $J = 7.4$ , 1 H) in the  $^1\text{H}$ -NMR spectrum and one anomeric C-atom at  $\delta(\text{C})$  105.7 in the  $^{13}\text{C}$ -NMR spectrum revealed that **2** possesses an additional  $\beta$ -linked sugar unit. The spectral data of the remaining resonances were almost the same as those of **1** [10][11]. The above data suggested that **2** is a 'tangshenoside-type' phenylpropanoid derivative [6]. The esterification of the 3-hydroxy-3-methylglutaric acid moiety was confirmed by the downfield shifted  $\text{CH}_2\text{O}$  *d* at  $\delta(\text{H})$  4.65 ( $\text{CH}_2(9''')$ ), and further supported by a HMBC cross-peak  $\text{CH}_2(9''')/\text{C}(5)$  (Fig. 2). The attachment of the  $\beta$ -glucopyranosyl moiety at C(4''') was established by an HMBC cross-peak  $\text{H}-\text{C}(1''')/\text{C}(4''')$ .

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### Experimental Part

*General.* Column chromatography (CC): silica gel (SiO<sub>2</sub>; 100–200 or 200–300 mesh; *Qingdao Ocean Chemical Co., Ltd.*, Qingdao, China), *ODS* (50 µm; *YMC*, Japan), and *Sephadex LH-20* (*Amersham Biosciences, GE Health Care*). Thin-layer chromatography (TLC): silica gel *GF<sub>254</sub>* plates (*Qingdao Ocean Chemical Co., Ltd.*, Qingdao, China). Optical rotation: *Perkin-Elmer 341* polarimeter; in MeOH at 22°. UV Spectra: *Shimadzu UV-240A* apparatus; λ<sub>max</sub> (log ε) in nm. IR Spectra: *Nicolet FT-IR-380* spectrometer; KBr pellets; in cm<sup>-1</sup>. <sup>1</sup>H- (500 MHz), <sup>13</sup>C- (125 MHz), and 2D-NMR (HMBC, HSQC) Spectra: *Bruker AV-500* spectrometer; in CD<sub>3</sub>OD; chemical shifts δ in ppm and coupling constants *J* in Hz. ESI-MS and HR-ESI-MS: *LCQ-Deca-XP<sup>plus</sup>* (*Thermo Finnigan*) and *Finnigan MAT-95* spectrometer, resp.; in *m/z*.

*Plant Material.* The roots of *C. tangshen* were collected in Chongqing, China, in January 2006, and identified by Prof. *Rui Peng* (Chongqing Academy of Chinese Materia Medica). A voucher (No. 06-01-28) specimen was deposited with the Herbarium of Chongqing Academy of Chinese Materia Medica.

*Extraction and Separation.* The air-dried, finely sliced roots of *C. tangshen* (25 kg) were extracted with 70% EtOH (3 × 120 l) under reflux for 2.0 h. After removal of the EtOH by evaporation, the resulting residue (7.5 kg) was suspended in H<sub>2</sub>O (8 l) and then partitioned successively with petroleum ether (1 × 4 l, 2 × 3 l), AcOEt (1 × 4 l, 2 × 3 l), and BuOH (1 × 4 l, 2 × 3 l). The BuOH fraction (1.5 kg) was dissolved in H<sub>2</sub>O, the soln. filtered, and the filtrate purified with the aid of a *D-101* macroporous resin column (20 × 80 cm), eluting successively with H<sub>2</sub>O, 30%, 50%, and 95% (v/v) EtOH: *Fractions 1–4*. *Fr. 2* (170 g) was firstly separated by CC (*ODS*, MeOH/H<sub>2</sub>O 1:9 → 4:6, then 100:0): *Fr. 2a–Fr. 2e*. *Fr. 2a* was subjected to CC (*ODS*, MeOH/H<sub>2</sub>O 2:8): **3** (30 mg). *Fr. 2b* was submitted to repeated CC (*Sephadex LH-20*, MeOH/H<sub>2</sub>O 1:1; then *RP-18* gel, MeOH/H<sub>2</sub>O 3:7): **1** (60 mg) and **4** (15 mg). *Fr. 3* (50 g) was subjected to CC (SiO<sub>2</sub>, gradient CHCl<sub>3</sub>/MeOH 80:1 → 1:1): *Fr. 3a–Fr. 3j*. *Fr. 3h* was subjected to CC (silica gel, CHCl<sub>3</sub>/MeOH 10:1 → 0:1); then *RP-18* gel (MeOH/H<sub>2</sub>O 4:6): **2** (22 mg).

3-[[6-O-[(2E)-3-(4-Hydroxyphenyl)-1-oxoprop-2-en-1-yl]-β-D-glucopyranosyl]oxy]-3-methylglutaric Acid (= *Tangshenoside V*; **1**): Colorless gum. [α]<sub>D</sub><sup>22</sup> = –3.40 (*c* = 0.65, MeOH). UV (MeOH): 311 (4.32). IR (KBr): 3432, 2924, 1706, 1634, 1604, 1174, 835. <sup>1</sup>H- and <sup>13</sup>C-NMR: *Table*. HR-ESI-MS (pos.): 493.1334 ([*M* + Na]<sup>+</sup>; calc. 493.1316).

3-[[6-O-[(2E)-3-(4-Hydroxyphenyl)-1-oxoprop-2-en-1-yl]-β-D-glucopyranosyl]oxy]-3-methylglutaric Acid (2E)-3-[4-(β-D-Glucopyranosyloxy)-3,5-dimethoxyphenyl]prop-2-en-1-yl Ester (= *Tangshenoside VI*; **2**): Colorless gum. [α]<sub>D</sub><sup>22</sup> = –10.2 (*c* = 0.10, MeOH). UV (MeOH): 227 (4.68), 277 (4.44), 309 (4.32). IR (KBr): 3432, 2927, 1706, 1633, 1604, 1172, 1126, 1072, 833. <sup>1</sup>H- and <sup>13</sup>C-NMR: *Table*. HR-ESI-MS (pos.): 847.2629 ([*M* + Na]<sup>+</sup>; calc. 847.2631).

### REFERENCES

- [1] Chinese Pharmacopoeia Commission, 'China Pharmacopoeia', Vol. 1, Chemical Industry Press, Beijing, 2005, p. 199.
- [2] Z.-T. Wang, Q. Du, G.-J. Xu, R.-J. Wang, D.-Z. Fu, T.-B. Ng, *Gen. Pharmacol.* **1997**, 28, 469.
- [3] Y.-P. Zhu, 'Chinese Materia Medica: Chemistry, Pharmacology, and Applications', Harwood Academic Publishers, Amsterdam, 1998, p. 158.
- [4] Z. T. Wang, T. B. Ng, H. W. Yeung, G. J. Xu, *Gen. Pharmacol.* **1996**, 27, 1347.
- [5] Q. He, E.-Y. Zhu, Z.-T. Wang, L.-S. Xu, Z.-B. Hu, *J. Chin. Pharm. Sci.* **2004**, 13, 212.
- [6] K. Mizutani, M. Yuda, O. Tanaka, Y.-I. Saruwatari, T. Fuwa, M.-R. Jia, Y.-K. Ling, X.-F. Pu, *Chem. Pharm. Bull.* **1988**, 36, 2689.
- [7] K. Mizutani, M. Yuda, O. Tanaka, Y.-I. Saruwatari, M.-R. Jia, Y.-K. Ling, X.-F. Pu, *Chem. Pharm. Bull.* **1988**, 36, 2726.
- [8] M. Yuda, K. Ohtani, K. Mizutani, R. Kasai, O. Tanaka, M.-R. Jia, Y.-R. Ling, X.-F. Pu, Y.-I. Saruwatari, *Phytochemistry* **1990**, 29, 1989.
- [9] Z.-T. Wang, G.-J. Xu, M. Hattori, T. Namba, *Shoyakugaku Zasshi* **1988**, 42, 339.
- [10] M. Cuendet, O. Potterat, K. Hostettmann, *Phytochemistry* **2001**, 56, 631.
- [11] W. G. Ma, R. X. Tan, N. Fuzzati, Q. S. Li, J.-L. Wolfender, K. Hostettmann, *Phytochemistry* **1997**, 45, 411.

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